

IACUC Semi-annual Core Facility Inspection Checklist

- **Purpose:** To provide the research laboratory staff with a checklist of items that will be inspected.
1. Procedure Areas
 - a. Space should be dedicated to animals use when animals are present in the lab.
 - b. Space should be free of drafts, windows, and laboratory traffic.
 - c. There should be access to biohazard waste containers (with an autoclavable liner) and a sharps container.
 - d. Procedure spaces should be clean and disinfected (best lab practice is before and after procedures).
 - e. Disinfectants should be within their expiration date. Acceptable disinfectants include:
 - ✓ Quaternary ammonium disinfectant wipes (e.g. – Santi Cloths)
 - ✓ 10% Bleach (if made fresh daily)
 - ✓ Chlorine dioxide (e.g. – Clidox or MB-10)
 - ✓ Glutaraldehydes (e.g. – Cetylcide or Cide Wipes)
 - ✓ Phenolics (e.g. – Lysol, TBQ, Note: should not be used if working with frogs)
 - ✓ Chlorhexidine (e.g. – Nolvasan or Hibiclens)
 - f. No animals from the university should be in the procedure space at the same time as animals from the VAMC. Procedure and facility space should be cleaned before and after animals from the VAMC are brought to the facility.
 - g. Eyewash stations should be flushed monthly and there should be a log kept by the lab documenting when the eye wash station was last flushed.
 2. Instruments and Equipment
 - a. Instruments/equipment should be cleaned and disinfected after use with each animal.
 - b. Instruments/equipment should be in good working order
 - c. Isoflurane vaporizers (if applicable)
 - ✓ Vaporizers should be calibrated and certified annually and be within their certification date.
 - ✓ Anesthetic gases should be scavenged.
 - ✓ If using scavenging canisters, the initial weight of the canister should be recorded (on the canister or on a log sheet kept by the canister) and the dates used and canister weight for that date should be recorded. The canisters should not exceed their expiration weight.
 3. Drugs and Reagents
 - a. All diluted drugs (e.g. – Buprenorphine, Ketamine/Xylazine, etc.) should be in sealed injectable vials, or alternatively, red-top blood collection tubes (if applicable). No injectable drugs or reagents should be in screw-top containers.

- b. All drugs and reagents should be labeled with and be within their expiration date (if the drug is diluted or a mix, then the expiration date is 6 months from the date it was mixed or if any component has an expiration date occurring prior to 6 months.)
- c. All drugs and reagents should be of pharmaceutical grade unless justified in your Animal Protocol.
- d. All controlled substances should be stored behind 2 locks at all times per DEA regulations and University of Iowa Guidelines (If applicable).

4. Record Keeping

- a. If you are anesthetizing animals with an injectable anesthetic for a non-surgical procedure, and the animal will recover from the anesthesia, then you must keep the following anesthesia records:
 - ✓ Date of surgery
 - ✓ PI Name
 - ✓ Animal Protocol number
 - ✓ Animal ID number
 - ✓ Species
 - ✓ Animal Weight
 - ✓ Anesthetic used, dose, and route
 - ✓ Time of induction of anesthesia
 - ✓ Time of recovery from anesthesia (awake and sternal) Monitoring right after surgery (animals should be continuously monitored until they are awake and sternal (upright). They then should be monitored every 15 minutes until ambulatory (no animals should be returned to OAR before they are able to walk).

Note: If your facility is located within an OAR vivarium, then you will need to follow all OAR guidelines and SOPs for cleaning and maintenance of the room(s). Please contact your facility supervisor for a checklist/log of tasks to complete and the frequency they must be done.